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(54) **CHLORINE DIOXIDE GAS FOR USE IN TREATING RESPIRATORY VIRUS INFECTION**

CHLORDIOXID GAS ZUR ANWENDUNG IN DER BEHANDLUNG VON ATMUNGSVIREN INFEKTIONEN

DIOXYDE DE CHLORE POUR L'UTILISATION DANS LE TRAITEMENT D'INFECTIONS CAUSÉES PAR VIRUS RESPIRATOIRES

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Description

TECHNICAL FIELD

5 [0001] The present invention relates to chlorine dioxide for use in treating respiratory virus infection.

BACKGROUND ART

10 [0002] In case e.g. a worker infected with a certain type of respiratory virus is present in an office or the like, there is the risk of infection with the same respiratory virus for the other works working within the same office. In such case, such infected workers may develop poor physical condition and experience eventual onset of the respiratory organ viral disease, thus leading to significant deterioration in the working efficiency in that office.

15 [0003] Here, the term respiratory virus is a generic name for various viruses which cause respiratory diseases such as pneumonia in animals. Among them are included influenza virus, parainfluenza virus, rhinovirus, bird flu virus, SARS virus, corona virus, etc. Infection with such respiratory virus and subsequent onset of respiratory virus disease can sometimes result in death of the subject.

[0004] Such respiratory viruses exist also in droplets exhaled from a respiratory organ of the infected subject. As the droplets can float in the air, inhalation of droplets containing the respiratory virus leads to spread of the infection and the spread of the respiratory virus.

20 [0005] Incidentally, though not commonly called "respiratory viruses", measles virus and rubella virus belong in the same category as above because inhalation by an uninfected subject of air contaminated with droplets from a respiratory organ of an infected subject causes secondary infection through the respiratory organ of the former.

25 [0006] For preventing spreading of respiratory virus, one effective measure is eliminating and/or deactivating the respiratory virus floating in the room (in the air). As one example of a method of deactivating respiratory virus floating in a room, there is a method called fumigation (see Non-Patent Document 1).

[0007] Non-Patent Document : "Sterilization/Disinfection Manual", Ishiyaku Publishing Inc. 1991, Sterilization/Disinfection Manual Compiling Committee.

30 [0008] JP 2002-272827A describes an infection prevention system for a treatment room comprising an air conditioning system into which chlorine dioxide is introduced.

DISCLOSURE OF THE INVENTION

OBJECT TO BE ACHIEVED BY INVENTION

35 [0009] Fumigation is a method wherein a hermetically closed room is filled with a gas prepared by evaporating an agent so as to kill insects, mites, fungus present inside the room. Some examples of the agent employed for fumigation are formalin, methyl bromide, aluminum phosphide, cyanide fume, etc.

40 [0010] However, such agents employed for fumigation are very poisonous for living bodies. So, during fumigation process, workers need to stop their works and evacuate from inside the room, thus leading to deterioration in the working efficiency.

[0011] The present invention has been made in view of the above-described state of the art.

MEANS TO ACHIEVE THE OBJECT

45 [0012] According to the invention, there is provided chlorine dioxide gas for use in the treatment of a disease caused by respiratory virus infection.

[0013] Chlorine dioxide gas has strong oxidative power and sterilizing power as well as strong virus deactivating power. For this reason, by exposing the respiratory virus to chlorine dioxide gas, the respiratory virus can be deactivated.

50 [0014] In this specification, the term "deactivation" refers to a condition of e.g. respiratory virus being destroyed, thus becoming unable to proliferate in a host, or a condition of the respiratory virus being still active, but hardly capable of proliferation in a host.

55 [0015] Therefore, if chlorine dioxide gas is supplied to a certain space and the concentration of the gas within the space is maintained at such a concentration as to deactivate the respiratory virus, even if respiratory virus is newly introduced into the space, this respiratory virus can be deactivated immediately. Accordingly, it is possible to prevent virus infection to the animal present within that space.

[0016] Moreover, as the concentration of chlorine dioxide within the space is such that the animal is allowed to live or stay, but the respiratory virus is deactivated, the animal can lead a normal living in the space. Therefore, it is possible to e.g. maintain air inside an office under the condition for deactivating respiratory virus, while allowing workers to

continue work in this office. Consequently, secondary infection with the virus can be effectively prevented to allow the workers to maintain their health. And, also, there occurs no deterioration in the working efficiency since the work need not be suspended during an air cleaning operation by e.g. fumigation.

5 [0017] Incidentally, the term "animal" refers to any animal or living creature which lives or stays within the space and can be infected with the respiratory virus, including mammals such as humans, domestic livestock, birds, reptiles, etc.

[0018] According to one embodiment of the present invention, chlorine dioxide may be supplied into a space where respiratory virus can be present by a gas concentration which allows the animal to live or stay, but can prevent onset of viral disease in the animal infected with the virus.

10 [0019] In this case, even if the animal is infected with the respiratory virus, the possibility of onset of viral disease in the infected animal can be reduced.

[0020] According to another embodiment of the present invention, chlorine dioxide may be supplied into a space where respiratory virus can be present by a gas concentration which allows the animal to live or stay, but can treat the animal infected with the virus and with onset of the viral disease associated therewith.

15 [0021] In the present case, even if the animal is infected with the respiratory virus and the animal experiences subsequent onset of the viral disease, the symptoms of this disease can be alleviated and its contraction period can be shortened.

[0022] According to a preferred embodiment of the present invention, the concentration of the chlorine dioxide gas within the space is set from 0.0001 ppm to 0.1 ppm.

20 [0023] With this characterizing construction, if the concentration of the chlorine dioxide gas within the space is under 0.0001 ppm, it is difficult to deactivate the respiratory virus. Further, as the working environment standard value for chlorine dioxide is 0.1 ppm, a value greater than 0.1 ppm can be harmful for the animal. Therefore, by setting the chlorine dioxide gas concentration within the space at the extremely low concentration from 0.0001 ppm to 0.1 ppm, the respiratory virus can be deactivated and at the same time the animal can live or stay more safely in the space.

25 [0024] Treatment of possible respiratory disease (e.g. pneumonia) which may develop after the infection can be effectively carried out.

[0025] According to a preferred embodiment of the present invention, the respiratory virus comprises influenza virus.

[0026] Treatment of various possible syndromes (e.g. fever attack, runny nose, sore throat, etc.) which may develop after the infection can be effectively carried out.

30 **BEST MODE OF EMBODYING THE INVENTION**

[EMBODIMENTS]

35 [0027] The present invention may be implemented with using an appropriate chlorine dioxide generator, for example.

[0028] Some non-limiting examples of chlorine dioxide generator suitable for the present invention include a large stationary apparatus, a small stationary apparatus, a small portable apparatus, and an instrument configured to charge a generating agent into gel or mix a dry agent therewith so as to generate a low concentration of chlorine dioxide gas for an extended period of time in a semi-automatic manner. However, the invention is not limited thereto, but it is also possible to employ an apparatus configured to discharge a low concentration of chlorine dioxide gas into a mouth.

40 [0029] The above chlorine dioxide generator apparatus comprises a reaction vessel, drug solution tanks capable of reserving respective drug solutions therein, liquid pumps, an air pump, a diluter, etc.

[0030] There are provided two kinds of drug solution tanks, each receiving therein chlorite water solution or acid therein. Some examples of chlorite usable are alkali metal chlorite (sodium chlorite, potassium chlorite, lithium chlorite), or alkali earth metal chlorite (calcium chlorite, magnesium chlorite, barium chlorite).. etc.. However, the invention is not limited thereto.

45 [0031] Some examples of acid usable are inorganic acids including hydrochloric acid, sulfuric acid, nitric acid, phosphoric acid. However, the invention is not limited thereto.

[0032] The liquid pump is provided as accompanying the drug solution tank. The pump is capable of precision control of liquid feed by means of a timer.

50 [0033] By the liquid pump, predefined amounts of chlorite water solution and acid solution are sent into the reaction vessel periodically, in which the solutions are mixed and reacted with each other to generate chlorine dioxide gas. The chlorine dioxide gas thus generated is mixed in the diluter with a predetermined rate of air sent by the air pump to be diluted to a predetermined concentration (preferably, 0.8 ppm).

55 [0034] The diluted chlorine dioxide gas is then supplied into a desired space where respiratory viruses can be present and the concentration of the chlorine dioxide gas within this space is maintained at a concentration which allows an animal to live or stay, but deactivates the viruses.

[0035] This concentration can be a concentration capable of preventing onset of a viral disease in the animal infected with the respiratory virus, or a concentration capable of treating the animal which has been infected with the respiratory

virus and in which the disease associated therewith has developed.

[0036] Specifically, based on results of influenza infection experiments using mice to be detailed later, this concentration is maintained to range from 0.05 ppm to 0.1 ppm.

5 [0037] Further, the concentration of chlorine dioxide can be set in the following manners for example. In the influenza infection experiment using mice to be described later, the mice were infected by administering thereto a high concentration of virus greater than 10 LD₅₀. In general, when there occurs respiratory viral infectious disease within such a large structure as a theater, an extremely low concentration of virus can be a problem. For instance, even such low concentration thereof as low as 0.02 LD₅₀ can cause infection.

10 [0038] In the above situation, the virus concentration in the air is about 1/500 of the above-noted concentration (10 LD₅₀/0.02 LD₅₀). Therefore, the concentration of chlorine dioxide gas required for preventing influenza infection is 0.0001 ppm (0.05/500).

15 [0039] That is to say, according to the present invention, by using the chlorine dioxide generator described above, the concentration of chlorine dioxide inside the space is maintained at the extremely low concentration (0.0001 to 0.1 ppm). With this, it becomes possible to maintain the chlorine dioxide gas concentration at such a concentration which can constantly be safe for the animal and which can, at the same time, deactivate the respiratory virus or deprive it of its infectability.

(Space)

20 [0040] The space to which the present invention is applicable includes a space that people enter/exit, such as a theater, an airport lobby, a business office, a classroom, or a space where animals are raised or plants are grown such as a mouse cage, a vinyl house, etc. or even such a "space" as oral cavity of a human, etc. However, the invention is not limited thereto. The invention can be applied to any other desired space which can assume, as desired, a closed state or an opened stage.

(Respiratory Virus)

25 [0041] Some examples of respiratory viruses include influenza virus, parainfluenza virus, rhinovirus, bird flu virus, SARS virus, corona virus, RS virus, etc. However, the invention is not limited thereto. The present invention is applicable to treatment of measles virus and rubella virus as inhalation by an un-infected subject of air contaminated with droplets from a respiratory organ of an infected subject causes secondary infection through the respiratory organ of the former, although these are not commonly called "respiratory viruses".

(Animal)

35 [0042] The living creature or organism to which the present invention is applied includes any animal or living creature which lives or stays within the space and can be infected with the respiratory virus, including mammals such as humans, domestic livestock, birds, reptiles, etc.

EXAMPLES

40 [0043] Next, the present invention will be described in details with reference to some examples thereof. It is understood; however, that the present invention is not limited thereto.

(Generation of Chlorine Dioxide Gas)

45 [0044] 0.25% sodium chlorite (NaClO₂) and 0.9% hydrochloric acid are charged and kept within separate drug solution tanks. Then, by using the liquid pump attached to each tank, each solution is sent to the reaction vessel.

50 [0045] Chlorine dioxide generated after the mixing and reaction between the sodium chloride and hydrochloric acid is added with air introduced thereto and the resultant product is discharged as a gas. In this, the discharged chlorine dioxide gas had a concentration of about 50 ppm. The discharged chlorine dioxide gas was then mixed in the diluter with air of a predetermined flow rate to be diluted to a low concentration.

55 [0046] In this experiment, the diluted and discharged chlorine dioxide gas had a concentration of 0.8 ppm. As will be described later, inside a mouse cage, the gas will be further diluted to 0.08 ppm or lower. The concentrations of chlorine dioxide gases were determined always by a chlorine dioxide meter (4330-SP, Interscan Corporation, U.S.A.).

(Preparation of Influenza Virus)

[0047] The influenza virus employed was prepared and obtained by proliferating Type A influenza strain A/PR8 (H1N1) in a culture medium containing 2% bovine fetal serum, with using, as a host, MDCK cell (ATCC CCL34) which is canine renal cell.

[0048] The viruses proliferated as above were suspended in a phosphate buffer solution (PBS) and then rendered into aerosol for subsequent use. In the above, the concentration of the suspended virus was adjusted to be 10, 100 or 1000 times of the concentration (LD₅₀) which, if introduced as aerosol into the mouse cage, would kill 50% of mice, respectively.

(Animal Experiment)

[0049] Animal experiment was conducted with using 8-week old male CD-1 mice. Fifteen mice, as one group, were put into a mouse cage dimensioned as: 25.5 cm x 36.8 cm x 8.0 cm. Into this, the aerosol of Type A influenza was introduced at the rate of 12.5 liter per minute.

[0050] In the above, at the same time, chlorine dioxide gas at 0.8 ppm concentration was fed at the flow rate of 0.6 to 1.8 liter per minute. The chlorine dioxide gas present within the mouse cage was diluted by the aerosol present therein.

[0051] In this experiment, concentration settings of 0 ppm, 0.03 ppm, 0.05 ppm and 0.08 ppm, as actually determined within the mouse cage, were possible for the chlorine dioxide gas. Data at these four kinds of concentration were obtained.

[0052] The exposure period for the mice to the virus aerosol was set to 15 minutes.

[0053] In principle, the introduction of chlorine dioxide gas was effected simultaneously with the introduction of the virus-containing aerosol. In some of the experiments, however, the introduction of chlorine dioxide gas was effected with delays relative to the introduction of the virus-containing aerosol. This was done in order to check whether the chlorine dioxide gas can prevent onset of viral disease resulting from the virus which has entered the lungs already.

[0054] As a control experiment, a phosphate buffer solution not containing any virus was employed and aerosol of this phosphate buffer solution was introduced into the mouse cage.

For those mice after the virus exposure, one mouse after another thereof was put into a separate mouse cage and kept therein in isolation for two weeks. During this period, in order to check presence/absence of virus infection and degree thereof (the number of virus), lung tissues were excised from five of the total fifteen mice.

[0055] The excised lungs were ground/crushed and then viruses were separated therefrom and the amount thereof was quantified.

[0056] For this quantification of the viruses within the lung, the crushed lung substance was suspended and its dilution series were prepared temporarily. Then, these were infected to the culture cells and TCID₅₀ values (50% tissue culture infection amount) were obtained.

[0057] On the other hand, as to the remaining 10 mice, life or death thereof was observed until day 14. During this period, if a mouse died in this period, its lung tissue was excised therefrom and fixed in formalin at that timing. If a mouse did not die in this period, then, its tissue was excreted and fixed in formalin on day 14. And, histopathologic analysis was conducted thereon by the standard method.

(Simultaneous Administration of 0.08 ppm Chlorine Dioxide Gas)

[0058] Mice were exposed to aerosol containing 1000 LD₅₀ virus for 15 minutes and to these mice, chlorine dioxide gas at 0.08 ppm concentration (final concentration inside the mouse cage) was administered simultaneously. The life and death of these mice are shown below (Table 1)

[Table 1]

Number of mice which died after exposure to virus (data for 10 mice in each group)							
days after exposure	3	4	5	8	9	10	14
0 ppm group	0	8	10	10	10	10	10
0.08ppm group	0	0	0	0	6	10	10

[0059] As may be apparent from Table 1 above, it may be understood that the deaths occurred with delay in the mice administered with the 0.08 ppm chlorine dioxide gas. On this difference, statistical significant difference test can be done. Its significance level (risk rate) was under 0.0001 (P< 0.0001). That is, this difference was found to have "statistical significance".

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[0060] Next, respecting the same experiment, from each 15 mice group 3 days after virus exposure, 5 mice were sampled from each group and their lung tissues were excreted and crushed and the virus contained therein was quantified (Table 2).

5

[Table 2]

Total virus amount in lung tissues on Day 3 (TCID ₅₀ values)					
serial mouse number	1	2	3	4	5
0 ppm group	10 ^{7.8}	10 ^{7.8}	10 ^{7.3}	10 ^{7.3}	10 ^{7.8}
serial mouse number	6	7	8	9	10
0.08 ppm group	10 ^{6.3}	10 ^{6.3}	10 ^{6.8}	10 ^{5.8}	10 ^{6.8}

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[0061] As shown in Table 2, when the numbers of virus in all the tissues were represented as TCID₅₀ values, the average value of the 0 ppm group was 10^{7.7}, and the average value of the 0.08 ppm administration group was 10^{6.4}, respectively. That is to say, the values of the group administered with the 0.08 ppm chlorine dioxide gas was only 5% of that of the control group (0 ppm group) (note the values are represented in the logarithm system)

20

[0062] Based on the above, it is believed that in the presence of chlorine dioxide gas, proliferation of virus was restricted or the amount of virus having infectability at the time of infection was small.

[0063] These conditions can be interpreted to be a condition where the respiratory viruses lost activity and could hardly proliferate in the bodies or mice or a condition where the infection to the mice was prevented due to insufficient infectability of the respiratory viruses.

25

[0064] From the above, it has been found that if influenza virus is present at the concentration of 1000 LD₅₀, inhibitory effect against onset of influenza can be obtained with administration of 0.08 ppm chlorine dioxide gas.

[0065] Subsequently to the above, a similar experiment was conducted with varying the concentration of virus to which mice were exposed.

30

[0066] In this experiment, aerosols of virus with concentrations of 10 LD₅₀ (10 times greater than LD₅₀) and 100 LD₅₀ (100 times greater than LD₅₀) were introduced to the mouse cages for 15 minutes; and at the same time, 0.08 ppm (final concentration inside the mouse cage) chlorine dioxide gas was introduced to the mouse cages.

[0067] As a result, in the case of the virus administered group with the 10 LD₅₀ virus concentration, no onset of influenza was found for 14 days in the group administered with 0.08 ppm chlorine dioxide. (Table 3).

[Table 3]

Number of mice which died after exposure to 10 LD ₅₀ virus (data for 10 mice in each group)							
days after exposure	3	4	5	8	9	10	14
0 ppm group	0	0	0	5	7	10	10
0.08 ppm group	0	0	0	0	0	0	0

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[0068] Also, the amount of virus in the lungs of the mice on Day 3 after the virus exposure was clearly lower for the chlorine dioxide administered group. (Table 4)

45

[Table 4]

Total virus amount in lung tissues on Day 3 after 10 LD ₅₀ virus exposure (TCID ₅₀ values)					
serial mouse number	21	22	23	24	25
0 ppm group	10 ⁵	10 ⁵	10 ^{5.3}	10 ^{5.1}	10 ⁵
serial mouse number	26	27	28	29	210
0.08 ppm group	10 ³	10 ^{3.2}	10 ^{3.3}	10 ³	10 ^{3.1}

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[0069] Further, in the virus administered group with 100 LD₅₀ virus concentration too, no onset of influenza after the virus exposure was found (Table 5) and the virus amount in the lung (Table 6) was small for the chlorine dioxide gas administered group.

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[Table 5]

Number of mice which died after exposure to 100 LD ₅₀ virus (data for 10 mice in each group)							
days after exposure	3	4	5	8	9	10	14
0 ppm group	0	0	3	6	8	10	10
0.08 ppm group	0	0	0	0	0	0	0

[Table 6]

Total virus amount in lung tissues on Day 3 after 100 LD ₅₀ virus exposure (TCID ₅₀ values)					
serial mouse number	31	32	33	34	35
0 ppm group	10 ^{6.3}	10 ^{6.2}	10 ^{6.1}	10 ^{6.1}	10 ^{6.6}
serial mouse number	36	37	38	39	310
0.08 ppm group	10 ^{5.5}	10 ^{5.8}	10 ^{5.6}	10 ^{5.7}	10 ^{5.9}

[0070] From the above, it has been found that in the cases of influenza virus being present at the concentrations of 10 LD₅₀ and 100 LD₅₀, sufficient inhibitory effect against onset of influenza can be obtained with administration of 0.08 ppm chlorine dioxide gas.

(Simultaneous Administration of 0.05 ppm Chlorine Dioxide)

[0071] Next, regarding the case of administration of 10 LD₅₀ concentration virus, investigation was made on the effect of simultaneous administration of an even lower concentration (0.05 ppm) of chlorine dioxide. The result showed that no onset of influenza was found in the group of chlorine dioxide administered group (Table 7) and the virus amount in the lung (Table 8) was small.

[0072] From the above, it has been found that in the cases of influenza virus being present at the concentrations of 10 LD₅₀, onset of influenza can be inhibited with simultaneous administration of 0.05 ppm chlorine dioxide gas.

[Table 7]

Number of mice which died after exposure to 10 LD ₅₀ virus with simultaneous administration of 0.05 ppm chlorine dioxide (data for 10 mice in each group)							
days after exposure	3	4	5	8	9	10	14
0 ppm group	0	0	4	5	8	10	10
0.05 ppm group	0	0	0	0	0	0	0

[Table 8]

Total virus amount in lung tissues on Day 3 after 10 LD ₅₀ virus exposure with simultaneous administration of 0.05 ppm chlorine dioxide (TCID ₅₀ values)					
serial mouse number	41	42	43	44	45
0 ppm group	10 ^{6.2}	10 ^{6.5}	10 ^{6.2}	10 ^{6.2}	10 ^{6.4}
serial mouse number	46	47	48	49	410
0.05 ppm group	10 ^{5.4}	10 ^{5.8}	10 ^{5.6}	10 ^{5.7}	10 ^{5.8}

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(Simultaneous Administration of 0.03 ppm Chlorine Dioxide)

[0073] Next, regarding the case of administration of 10 LD₅₀ concentration virus, investigation was made on the effect of simultaneous administration with further reducing the concentration of chlorine dioxide to 0.03 ppm. The result showed that between the group of chlorine dioxide administered group (0.03 ppm) and the control group (0 ppm) no significant difference was found in the number of onsets of influenza (Table 9) or the virus amount in the lung (Table 10).

[Table 9]

Number of mice which died after exposure to 10 LD ₅₀ virus with simultaneous administration of 0.03 ppm chlorine dioxide (data for 10 mice in each group)							
days after exposure	3	4	5	8	9	10	14
0 ppm group	0	0	3	7	7	10	10
0.03 ppm group	0	0	3	8	9	10	10

[Table 10]

Total virus amount in lung tissues on Day 3 after 10 LD ₅₀ virus exposure with simultaneous administration of 0.03 ppm chlorine dioxide (TCID ₅₀ values)					
serial mouse number	51	52	53	54	55
0 ppm group	10 ^{6.1}	10 ^{6.4}	10 ^{6.0}	10 ^{6.1}	10 ^{6.2}
serial mouse number	56	57	58	59	510
0.03 ppm group	10 ^{6.3}	10 ^{6.3}	10 ^{6.1}	10 ^{6.9}	10 ^{6.3}

[0074] From the above, it has been found that in the cases of influenza virus being present at the concentration of 10 LD₅₀, simultaneous administration of 0.03 ppm chlorine dioxide gas is not effective respecting inhibition of influenza in mice.

[0075] Hence, it was confirmed that as the influenza onset inhibitory effect in the case of simultaneous administration of chlorine dioxide gas, the chlorine dioxide gas concentration of at least 0.05 ppm or more is effective in case the virus concentration is 10 LD₅₀ amount.

[0076] Based on the above-described experiments, the chlorine dioxide gas concentration to be maintained at a space can be set in a manner as follows for instance.

[0077] In general, when there occurs respiratory viral infectious disease within such a large structure as a theater, an extremely low concentration of virus can be a problem. For instance, even such low concentration as low as 0.02 LD₅₀ can cause infection.

[0078] In the above situation, the virus concentration in the air is about 1/500 of the above-noted concentration (10 LD₅₀/ 0.02 LD₅₀). It follows that the chlorine dioxide concentration required for preventing influenza virus infection is 0.0001 ppm (0.05/500). Therefore, the effect of deactivating virus and preventing infection can be expected with setting of the chlorine dioxide concentration to such low concentration values as above.

(Delayed Administration of 0.08 ppm Chlorine Dioxide Gas)

[0079] Next, regarding the 10 LD₅₀ concentration virus administered group, the effect of chlorine dioxide gas was investigated for cases of administering 0.08 ppm chlorine dioxide gas with delays of 5 minutes, 10 minutes and 15 minutes respectively from the start of virus exposure. In the respective cases, the exposure period to the chlorine dioxide gas was 15 minutes.

[0080] The object of this experiment is to confirm whether chlorine dioxide gas when administered with a delay is effective for preventing onset of influenza against viruses which have already reached the lungs and proliferated there, that is, whether it has a "treating effect" for influenza or not.

[0081] The results are shown in Table 11 and Table 12 below.

[Table 11]

Number of mice which died after exposure to 10 LD ₅₀ virus with delayed administration of 0.08 ppm chlorine dioxide (data for 10 mice in each group)							
Days after exposure	3	4	5	8	9	10	14
5 minutes delayed administration group	0	0	3	7	7	10	10
10 minutes delayed administration group	0	0	4	9	10	10	10
15 minutes delayed administration group	0	2	6	10	10	10	10

[Table 12]

Total virus amount in lung tissues on Day 3 after 10 LD ₅₀ virus exposure with delayed administration of 0.08 ppm chlorine dioxide (TCID ₅₀ values)					
Serial Mouse Number	61	62	63	64	65
5 minutes delayed administration group	10 ^{6.1}	10 ^{6.4}	10 ^{6.0}	10 ^{6.1}	10 ^{6.2}
Serial Mouse Number	71	72	73	74	75
10 minutes delayed administration group	10 ^{6.3}	10 ^{6.3}	10 ^{6.1}	10 ^{6.9}	10 ^{6.3}
Serial Mouse Number	81	82	83	84	85
15 minutes delayed administration group	10 ^{6.8}	10 ^{6.7}	10 ^{7.3}	10 ^{7.7}	10 ^{7.7}

[0082] The results of determination of the total virus amounts showed that the amounts were smaller than expected in spite of the fact that chlorine dioxide gas was administered after lapse of 15 minutes from the virus exposure. Based on this, it is reasoned that delayed administration of chlorine dioxide gas can alleviate the symptoms of influenza.

[0083] This result shows that even when influenza virus has already been fixed to the tissue of respiratory organ and proliferated, symptoms of influenza can be restricted with delayed administration of chlorine dioxide gas. This suggests the potential of chlorine dioxide gas achieving treating effect to animals with onset of influenza.

(Experiment of Protein Denaturation with Low Concentration Chlorine Dioxide)

[0084] In this embodiment, it has been described based on influenza virus infection experiments using mice that chlorine dioxide gas, even at an extremely low concentration of e.g. 0.05 to 0.1 ppm, is capable of deactivating virus or depriving its infectability.

[0085] Such anti-virus effect of chlorine dioxide can be explained based on the following experiment.

[0086] In this experiment, glucose-6-phosphate dehydrogenase (referred to shortly as "G6PD" hereinafter) which is an enzyme of yeast was reacted with a low concentration of chlorine dioxide and investigation was made to see how the enzyme activity of G6PD would change.

[0087] Chlorine dioxide was dissolved in PBS buffer (20 mM sodium phosphate buffer, pH7, 130 mM NaCl) to final concentrations of 0.1, 1, 10, 100, 1000 μM (0.007, 0.07, 0.7, 7, 70 ppm, respectively if converted into ppm concentration values) (these will be referred to as chlorine dioxide solutions hereinafter). G6PD was dissolved in each chlorine dioxide solution to a final concentration of 80 μg/mL and reaction was allowed to continue at 25°C for 2 minutes.

[0088] The enzyme activity of G6PD was determined by observing absorption of NADPH by a spectral photometer, with using NADP and glucose-phosphoric acid as the substrate. Specific reaction conditions were set according to the manufacturer's (Sigma Corp.) instructions.

[0089] The determination results of the enzyme activity are shown in Fig. 1. The concentration values on the horizontal axis are shown as being converted into the unit of ppm. The "specific activity" on the vertical axis represents enzyme

activity (unit/mg) per 1 mg of protein

[0090] The results show that when the chlorine dioxide concentration was 0.07 ppm, there was about 60% reduction (45/71) in the enzyme activity, as compared with the case of chlorine dioxide concentration being 0 ppm.

[0091] Therefore, it was found that even a very low concentration of chlorine dioxide as low as about 0.07 ppm can inhibit enzyme activity. This suggests that chlorine dioxide, even at an extremely low concentration range as proposed by the present invention, provides the anti-virus effect of inhibiting enzyme activity.

(Experiment for Confirming Effectiveness of Chlorine Dioxide Against Influenza Virus)

[0092] Next, there will be described an experiment conducted to investigate the effect of denaturation of surface proteins present in influenza virus in order to confirm the effectiveness of chlorine dioxide against influenza virus.

[0093] On the surface of influenza virus particle, there are two kinds of protein, referred to as hemagglutinin (referred to as HA hereinafter) and neuraminidase (referred to as NA hereinafter).

[0094] HA is a protein required at the initial stage of virus infection, that is, required for the virus to bind to the host cell surface and this protein has a function of promoting virus infection. On the other hand, NA is a protein which provides the function of cutting bond between offspring virus proliferated in the host cell and the surface of the host cell, thus separating the virus from the host cell surface. This facilitates diffusion of offspring viruses and spreading of virus infection.

[0095] Therefore, with deletion of the HA function, the virus becomes unable to infect or deletion of the NA function can minimize the number of host cells which will be killed by infection. Therefore, it is believed that the infectability of virus will be reduced when at least one of these two proteins loses its function.

[0096] Then, investigation was made to see how the functions of the two proteins would change when influenza virus was reacted with chlorine dioxide.

[0097] In this experiment, in order to clearly study the denaturing effect of the two proteins relative to chlorine dioxide, the concentration of chlorine dioxide was set so as to range from 2.7 to 21.4 ppm.

[0098] Chlorine dioxide was dissolved in PBS buffer (20 mM sodium phosphate buffer to final concentrations of 40, 80, 160, 240 and 320 μM (2.7, 5.4, 10.7, 16.1 and 21.4 ppm, respectively if converted into ppm concentration values) (these will be referred to as chlorine dioxide solutions hereinafter). Influenza virus was dissolved in each chlorine dioxide solution to a final concentration of 77 μg/mL and reaction was conducted on ice for 2 minutes.

[0099] After the reaction, titer values of HA were determined by a known blood coagulation reaction test. The results of determination are as shown in Table 13.

[Table 13]

Concentration of reacted chlorine dioxide (ppm)	0	2.7	5.4	10.7	16.1	21.4
HA titer value	512	256	32	32	16	4

[0100] Further, regarding the case of chlorine dioxide concentration being 5.4 ppm, investigation was made on variation in titer value according to the reaction period. The results of determinations are shown in Table 14.

[Table 14]

Reaction period (seconds)	0	5	10	20	30	40	60	120
HA titer value	256	16	16	16	16	16	16	16

[0101] This suggested that when influenza virus is reacted with chlorine dioxide, HA is denatured immediately to lose its function.

[0102] Table 15 shows the determination results of NA titer values.

[Table 15]

Concentration of reacted chlorine dioxide (ppm)	0	2.7	5.4	10.7	16.1	21.4
NA titer value	38.8548	37.3824	32.7322	18.7712	10.4366	7.2408
standard deviation	0.698112	1.59382	1.095355	0.677734	0.265636	0.118774

[0103] This suggested that when influenza virus is reacted with chlorine dioxide, NA is denatured immediately to lose

its function.

[0104] Based on the above, it was confirmed that chlorine dioxide reduces the infectability of influenza virus.

[Other Embodiments]

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[0105] In the above embodiment, as an example of the "space", a place where virus is present, such a location as a theater where humans enter/exit was described. However, the present invention is not limited thereto. A location where humans enter/exit can be in a liquid such as a swimming pool, bath, etc.

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[0106] Further, in the foregoing, animals were described as living creatures as the subjects. However, the present invention is not limited thereto. The invention can be applied to plants which have the possibility of being infected with respiratory virus.

INDUSTRIAL APPLICABILITY

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[0107] The present invention can be used in treating respiratory virus infection.

BRIEF DESCRIPTION OF DRAWING

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[0108] [Fig. 1] a view showing determination of enzyme activity of G6PD reacted with chlorine dioxide.

Claims

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1. Chlorine dioxide gas for use in the treatment of a disease caused by the infection of a respiratory virus.
2. The chlorine dioxide gas for use according to claim 1, wherein the respiratory virus is influenza virus.
3. The chlorine dioxide gas for use according to claim 1 or claim 2, wherein the concentration of chlorine dioxide ranges from 0.0001 ppm to 0.1 ppm.

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Patentansprüche

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1. Chlordioxidgas zur Verwendung bei der Behandlung einer durch Infektion mit einem Atemwegsvirus hervorgerufenen Erkrankung.
2. Chlordioxidgas zur Verwendung nach Anspruch 1, wobei es sich bei dem Atemwegsvirus um ein Grippe-virus handelt.
3. Chlordioxidgas zur Verwendung nach Anspruch 1 oder Anspruch 2, wobei die Chlordioxidkonzentration im Bereich von 0,0001 ppm bis 0,1 ppm liegt.

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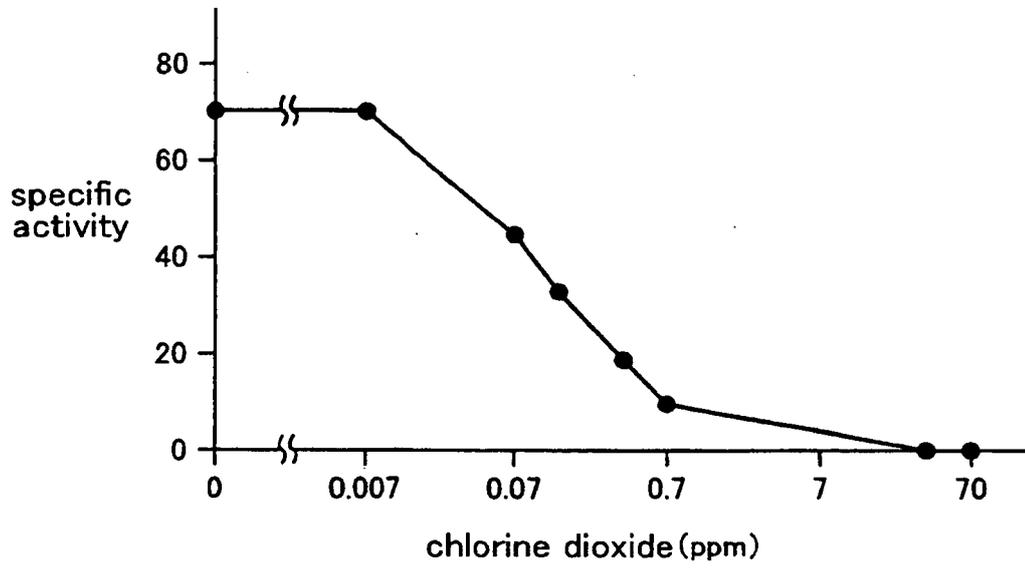
Revendications

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1. Dioxyde de chlore gazeux pour une utilisation dans le traitement d'une maladie provoquée par l'infection par un virus respiratoire.
2. Dioxyde de chlore gazeux pour une utilisation selon la revendication 1, **caractérisé en ce que** le virus respiratoire est l'influenzavirus.
3. Dioxyde de chlore gazeux pour une utilisation selon la revendication 1 ou la revendication 2, **caractérisé en ce que** la concentration en dioxyde de chlore va de 0,0001 ppm à 0,1 ppm.

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FIG.1



REFERENCES CITED IN THE DESCRIPTION

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Patent documents cited in the description

- JP 2002272827 A [0008]

Non-patent literature cited in the description

- Sterilization/Disinfection Manual. Ishiyaku Publishing Inc, 1991 [0007]